

胀果甘草悬浮培养细胞合成甘草总黄酮

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摘要：比较了胀果甘草 (*Glycyrrhiza inflata*) 悬浮细胞在逐级放大摇瓶中的生长、黄酮产量以及营养消耗过程，以便了解其放大规律。结果表明，在 250 和 500 mL 摇瓶中，细胞的最大生物量、黄酮产量以及最大比生长速率没有显著性差异，但是在 1 L 的摇瓶中，这三种参数都显著地降低，分别比 250 mL 摇瓶中降低了 27%，30% 和 27%。在逐级放大的摇瓶中，氮、磷、铵浓度都随着培养时间延长而逐渐降低，尽管在 1 L 的摇瓶中磷消耗得最慢，但三种摇瓶中磷在细胞生长对数期基本都被消耗尽了。此外，硝态氮在第 18 天时基本被消耗完，而铵态氮在细胞收获时仍能维持在 100 mg/L。因此在反应器中培养时，主要的培养条件还需进一步优化。

关键词：胀果甘草细胞；生长；黄酮产量；营养消耗；放大培养

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Production of Flavonoids in Cell Suspension Culture of
Glycyrrhiza inflata (Leguminosae)

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Abstract: The cell biomass, flavonoids production and nutrients consumption were monitored to characterize the culture of progressive scale-up of *Glycyrrhiza inflata* . The maximum biomass, flavonoids production and the maximum specific growth rate in the culture of 250 and 500 mL flasks remain similar, but were significantly higher than that of 1 000 mL flask . The three parameters in the culture of 1 000 mL flask were 27% , 30% , and 27% lower than that of 250 mL flask , respectively . The concentrates of phosphate, nitrate and ammonium in progressive scale-up flasks decreased similarly . Phosphate and nitrate were almost exhausted on day 10 and 18 in all flasks, respectively; while ammonium maintained about 100 mg/L till cells were harvested . The basic culture conditions needed to be further optimized for higher flavonoids production on a bio-reactor scale .

Key words: *Glycyrrhiza inflata*; Growth; Flavonoids; Nutrients consumption; Scale-up flask

Glycyrrhiza inflata Bat . is a traditional Chinese medicine . One of the major compounds, flavonoids have significant anti-oxidation activity, antitumor activity (Kanazawa *et al* . 2003), antihuman immunodeficiency virus activity (Hatano *et al* . 1988) and anti-

bacterial activity (Fukai *et al* . 2002) .

Thus , the *G. inflata* plant has fallen short of supply with an increasing demand for flavonoids . Plant cell culture is a useful method for the production of valuable secondary metabolites .

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However, there were few studies on cell suspension culture of *G. inflata*, and its nutrients consumption has not yet been systematically investigated. Cell growth and flavonoids biosynthesis are sensitive to environmental conditions. Furthermore, scaling up the culture from flask to bioreactor is very difficult. Consequently, in the present study, the cell growth, flavonoids accumulation and nutrient consumption were investigated in scale-up flasks to understand the characteristic of *G. inflata* cell, which may be beneficial for scaling up in the bioreactor.

Materials and Methods

G. inflata cell were maintained in Murashige and Skoog (MS) culture medium containing 3% sucrose supplemented with 2,4-dichlorophen oxyacetic acid (2,4-D, 0.5 mg/L), naphthalene acetic acid (NAA, 0.5 mg/L) and 6-benzyladenine (6-BA, 0.5 mg/L). For experiments, cells were cultured at 25 °C on a rotatory shaker (120 r/min) with 5% inoculum in 250, 500 mL and 1 L flasks with 70, 140 and 280 mL medium (pH 5.8), respectively. The cells were harvested by filtration, and dried at 50 °C to constant dry weight (DW). The maximum specific growth rates were calculated by the formula: $(X_2 - X_1) / (t_2 - t_1)$, where X_2 and X_1 were the DW of the cell at t_2 and t_1 days during the exponential growth phase, respectively.

Nitrate in the medium was determined according to Hecht & Mohr (1990). Salicylic acid- H_2SO_4 (5%) was put into the medium, 20 min later, 8% NaOH was added in. Then the absorbency was determined at 410 nm at 25 °C. Ammonium was assayed as reported by Moore & Stein (1948). After 2 mL dilute medium, 3 mL ninhydrin reagent and 0.1 mL 1% ascorbic acid were incubated together in 100 °C water for 15 min, and slowly cooled in air. The absorbency was determined at 580 nm. Additionally, phosphate was determined by colorimetry at 660 nm when

1 mL dilute medium and 3 mL phosphate reagent (2.5% ammonium molybdate 10% ascorbic acid water = 1:1:2) was placed in 45 °C water for 20 min (Li, 1998).

The flavonoids were extracted with 30 volumes of ethanol/water (70/30, v/v) by sonication for 1 h at 25 °C. After centrifugation at 10 000 rpm for 6 min, the supernatant was extracted three times with EtOAc, then with 95% ethanol. The combination of flavonoids in cells and medium was determined by colorimetry according to Zhang *et al.* (2001). Rutin was used as a standard sample.

Results and Discussion

Cultured with a cycle of 22 days, the maximum biomass, flavonoids production and the maximum specific growth rate of the cells all decreased with progressive scale-up flasks, and by 27%, 30% and 27% in 1 L flasks compared to that in 250 mL flasks, respectively (Table 1). But the differences were insignificant between 250 and 500 mL flasks ($P > 0.05$), which was agreed with the observation in *Catharanthus roseus* cell suspension culture (Zheng *et al.* 1998). Additionally, a little foam was observed on the surface of the

Table 1 Compare of the maximum biomass, flavonoids production and maximum specific growth rate in cell suspension culture of *G. inflata* in progressive scale-up flasks. Data with different superscript letters are significantly different (one-way ANOVA followed by a multiple comparison; a, b: $P < 0.05$; B: $P < 0.01$, $n = 3$).

Flasks	Maximum biomass (g DW/L)	Flavonoids production (mg/L)	Maximum specific growth rate (g DW/L/d)
250 mL	14.5 ± 0.61 ^a	80.35 ± 4.2 ^a	0.66 ± 0.03 ^a
500 mL	13.3 ± 0.82 ^a	73.63 ± 4.1 ^a	0.61 ± 0.04 ^a
1 L	10.6 ± 0.75 ^B	56.45 ± 1.4 ^B	0.48 ± 0.04 ^b

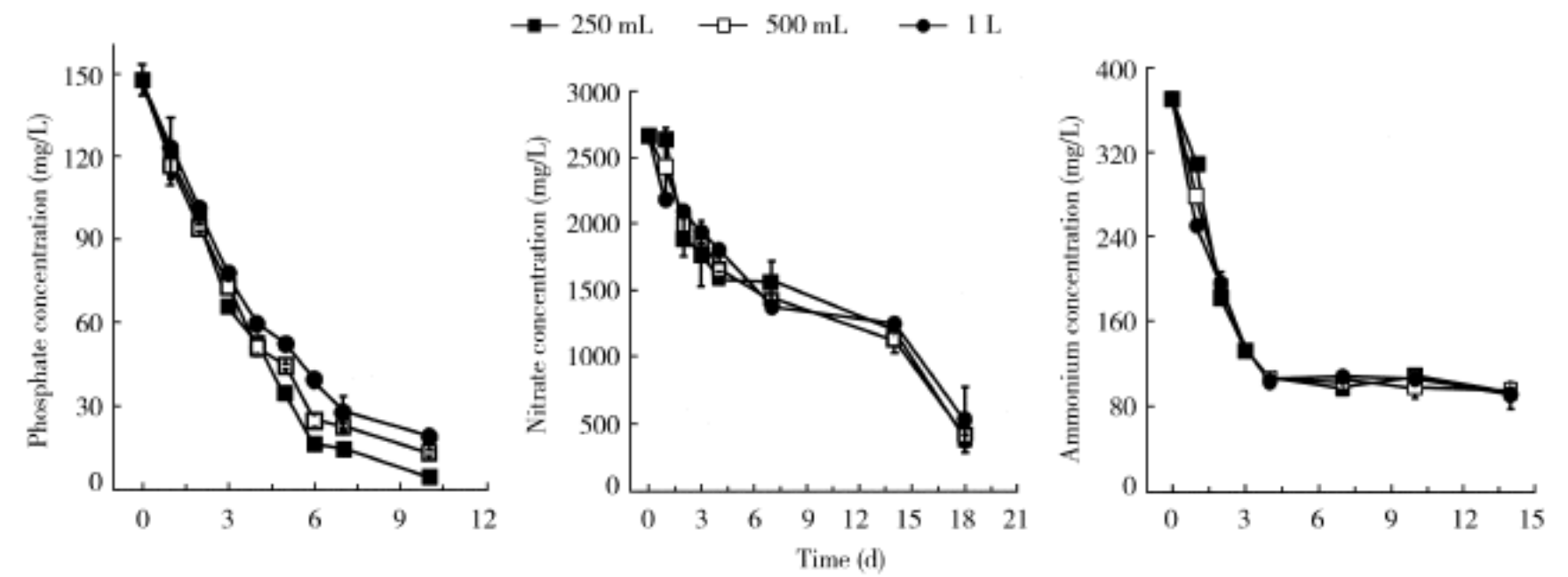


Fig. 1 Time courses of phosphate, nitrate and ammonium consumption in cell suspension culture of *G. inflata* grown in MS medium, and represent for 250, 500 mL and 1 L flasks, respectively. Each data indicated the means of three independent experiments.

1 L flasks in which the cells appeared brown and less healthy in the later stage of cultivation. Zheng *et al.* (1998) found that the oxygen mass transfer ($K_L a$) in 10 L bioreactor was significantly higher than that in flasks. Ten Hoopen (1994) reported that shear stress was the main reason making the cells brown. Thus, the decreased biomass of brown cell and flavonoids production in 1 L flasks might be due to shear stress and the decreased dissolved oxygen.

The trends of phosphate, nitrate and ammonium consumption in progressive scale-up flasks were very similar that the nutrients all decreased sharply (Fig. 1). At the initial several days, the phosphate consumptions in 250, 500 mL and 1 L flasks were almost alike, but later it was the slowest in 1 L flasks. The phosphate in all flasks was almost exhausted on day 10 at the logarithm phase, which was observed in many plants cell cultures (Shin *et al.* 2003). Most plant cells utilize ammonium firstly and nitrate later (Shin *et al.* 2003). In the study, the ammonium consumption was faster than nitrate at the beginning of the cultivation. Low concentrations of ammonium stimulate nitrate uptake and of nitrate inhibit ammonium uptake (Dortch, 1990). Thus, the nitrate was almost run out of on day 18, while the ammonium still maintained about 100 mg/L when the cells were harvested. Consequently, the main culture conditions needed to be further optimized for higher flavonoids production on a bioreactor scale.

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